Test for the direct qualitative detection of SARS-CoV-2, Influenza A, B and RSV on a four-channel qPCR cycler



Read the entire Instructions for Use (IFU) and follow them carefully before performing the test. Deviations from the given test protocol can lead to incorrect results. Good laboratory practice should be followed during the test.

Intended Pupose

The res4plex direct RT-PCR test is an assay for in vitro examination of viral RNA in nasal, nasopharyngeal or oral/oropharyngeal swabs to provide information to aid to diagnose patients under suspicion of respiratory diseases: SARS-CoV-2, influenza A, influenza B, and respiratory syncytial virus A/B (RSV A/B). The IVD medical device detects the RNA of the aforementioned pathogens by gualitative measurements based on RT-qPCR and is intended for use in medical laboratories or health institutions by laboratory personnel specifically trained in RT-qPCR and in vitro diagnostic techniques. It has to be used in combination with conventional nucleic acid extraction systems for RNA extraction and RT-qPCR cyclers for detection and analysis.

Package Content for 96 reactions

Solution A (green; 1.1 mL), Internal Control (blue; 400 µL) and Positive Control (red; 20 µL); Quick Start Protocol.

Notes before starting

The starting material for the res4plex direct RT-PCR test is 10 μ L/reaction RNA isolated from biological specimens (respiratory samples). Appropriate RNA extraction needs to be conducted according to the manufacturer's instructions. RNA extraction reagents are not part of the res4plex direct RT-PCR test. One Positive and one Negative Control should be included in each PCR run.

Material provided by user

Channel settings for FBC107-Cx

- RNA isolation kit
- Negative Control
- Adequate pipettes and sterile filter-tips for PCR testing (DNase/RNase-free)
- qPCR microtiter plate or reaction tubes; table centrifuge
- qPCR instrument (any four-channel qPCR cycler)

<u>Test procedure</u>

RNA Extraction

- Thaw all reagents completely.
- 2. Add Internal Control to the RNA preparation process in accordance to the laboratory's standard procedure (e.g., add 4µL/sample to lysis buffer).
- 3. Perform RNA extraction according to your laboratory's standard procedure.

RT-PCR

- Pipette 10 µL/well of solution A into the PCR microtiter plate/reaction tubes. 4.
- Add 10 µL/well of eluate from RNA extraction; add 10 µL Positive Control per run; add 10 µL Negative Control per run. 5.
- Close the microtiter plate with an adhesive optical film or the reaction tubes with the lids provided. 6.
- Briefly centrifuge the microtiter plate/reaction tubes. 7.
- 8. Place the filled microtiter plate/reaction tubes in the qPCR cycler. Start program.

Instrument settings

Steps	Temperature [°C]	Time	Number of cycles		CoV2_N/E	IC	RSV	Flu /	
Reverse transcription	55	10 min	1x	Reporter dye	FAM	HEX	Red 610	Су	
Initial denaturation	95	2 min	1x	Colour	green	yellow-green	orange	re	
Denaturation	95	5 sec	45	Emission [nm]	520	560	610	67	
Amplification/Elongation	61	25 sec	45x	Quencher	Black Hole Quencher				

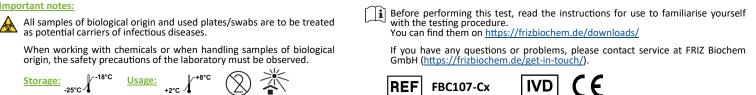
Interpretation of test results

Positive samples (+) show a qPCR typical amplification curve that crosses a certain threshold generating the Ct value. Co-infections of two or more pathogens are possible, but with low probability of occurrence and are therefore not included in the table below.

The results are used to identify SARS-CoV-2, Influenza A, Influenza B and RSV RNA. Positive results are an indication of the presence of the respective virus(es). A negative result does not rule out the presence of the respective pathogen, as the results depend on correct sampling and a sufficient amount of RNA to be detected.

CoV2_N/E	RSV	Flu A/B	IC			6800	 	********	Sample
FAM	Red610	Cy5	HEX	Result	Interpretation	5800	 	********	positive (+)
+	-	-	+/-	Valid	SARS-CoV-2 detected.	4800	 		
-	+	-	+/-	Valid	RSV detected.	B 3800			
-	-	+	+/-	Valid	Influenza A/B detected.	2800	+		
-	-	-	+	Valid	No SARS-CoV-2, Influenza A/B or RSV detected.	1800 800	<u></u>		threshold
-	-	-	-	Invalid	The test result can not be evaluated.	-200	20 25 30	+ + + + + + + + + + + + + + + + + + + +	Sample

Important notes:



FRIZ Biochem GmbH * Floriansbogen 2-4 * D-82061 Neuried * Germany * Tel +49 (0) 89 - 72 44 09 25 * Fax +49 (0) 89 - 72 44 09 10 info@frizbiochem.de • www.frizbiochem.de

Cycle number