FBC107-Ax Quick Start Protocol

res4plex direct RT-PCR

Test for the direct qualitative detection of SARS-CoV-2, Influenza A, B and RSV on a Light Cycler 480 II or equiv.



Read the entire Instructions for Use (IFU) and follow them carefully before performing the test. Deviations from the given test protocol can lead to incorrect results. Good laboratory practice should be followed during the test.

Intended Pupose

The res4plex direct RT-PCR test is an assay for in vitro examination of viral RNA in nasal, nasopharyngeal or oral/oropharyngeal swabs to provide information to aid to diagnose patients under suspicion of respiratory diseases: SARS-CoV-2, influenza A, influenza B, and respiratory syncytial virus A/B (RSV A/B). The IVD medical device detects the RNA of the aforementioned pathogens by qualitative measurements based on RT-qPCR and is intended for use in medical laboratories or health institutions by laboratory personnel specifically trained in RT-qPCR and in vitro diagnostic techniques. It has to be used in combination with conventional nucleic acid extraction systems for RNA extraction and RT-qPCR cyclers for detection and analysis.

Package Content for 96 reactions

Solution A (green; 1.1 mL), Internal Control (blue; 400 µL) and Positive Control (red; 20 μL); Quick Start Protocol.

Notes before starting

The starting material for the res4plex direct RT-PCR test is 10 μ L/reaction RNA isolated from biological specimens (respiratory samples). Appropriate RNA extraction needs to be conducted according to the manufacturer's instructions. RNA extraction reagents are not part of the res4plex direct RT-PCR test. One Positive and one Negative Control should be included in each PCR run.

Material provided by user

- RNA isolation kit
- Negative Control
- Adequate pipettes and sterile filter-tips for PCR testing (DNase/RNase-free)
- · qPCR microtiter plate or reaction tubes; table centrifuge
- qPCR instrument (e.g. Light Cycler 480 II or equiv.)

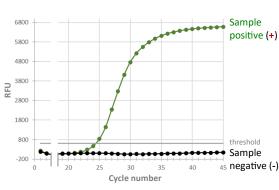
Test procedure V **RNA Extraction** Thaw all reagents completely. Add Internal Control to the RNA preparation process in accordance to the laboratory's standard procedure (e.g., add 4µL/sample to lysis buffer). 3. Perform RNA extraction according to your laboratory's standard procedure. RT-PCR Pipette 10 µL/well of solution A into the PCR microtiter plate/reaction tubes. 4. Add 10 μL/well of eluate from RNA extraction; add 10 μL Positive Control per run; add 10 μL Negative Control per run. Close the microtiter plate with an adhesive optical film or the reaction tubes with the lids provided. Briefly centrifuge the microtiter plate/reaction tubes. 7. Place the filled microtiter plate/reaction tubes in the qPCR cycler. Start program. **Instrument settings Channel settings for FBC107-Ax** Temperature [°C] Time Number of cycles Steps IC CoV2_N CoV2_E Flu B RSV Flu A Reverse transcription 55 10 min 1x Reporter dye Cyan500 **FAM** HEX Red 610 Cy5 Initial denaturation 95 2 min 1x Colour cyan green yellow-green orange red Denaturation 95 5 sec Emission [nm] 480 670 520 560 610 45x Amplification/Elongation 61 25 sec Black Hole Quencher Quencher

Interpretation of test results

Positive samples (+) show a qPCR typical amplification curve that crosses a certain threshold generating the Ct value. Co-infections of two or more pathogens are possible, but with low probability of occurrence and are therefore not included in the table below.

The results are used to identify SARS-CoV-2, Influenza A, Influenza B and RSV RNA. Positive results are an indication of the presence of the respective virus(es). A negative result does not rule out the presence of the respective pathogen, as the results depend on correct sampling and a sufficient amount of RNA to be detected.

CoV2_N/E	Flu B	RSV	Flu A	IC		
FAM	HEX	Red 610	Cy5	Cyan 500	Result	Interpretation
+	-	-	-	+/-	Valid	SARS-CoV-2 detected.
-	+	-	-	+/-	Valid	Influenza B detected.
-	-	+	-	+/-	Valid	RSV detected.
-	-	-	+	+/-	Valid	Influenza A detected.
-	-	-	-	+	Valid	No SARS-CoV-2, Influenza A, Influenza B or RSV detected.
-	-	-	-	-	Invalid	The test result can not be evaluated.



Important notes:

Storage:



All samples of biological origin and used plates/swabs are to be treated as potential carriers of infectious diseases.

When working with chemicals or when handling samples of biological origin, the safety precautions of the laboratory must be observed.

Usage:





Before performing this test, read the instructions for use to familiarise yourself with the testing procedure. You can find them on https://frizbiochem.de/downloads/

If you have any questions or problems, please contact service at FRIZ Biochem GmbH (https://frizbiochem.de/get-in-touch/).



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